

Bioorganic & Medicinal Chemistry Letters 11 (2001) 265-270

Antagonists of the Human CCR5 Receptor as Anti-HIV-1 Agents. Part 2: Structure—Activity Relationships for Substituted 2-Aryl-1-[N-(methyl)-N-(phenylsulfonyl)amino]-4-(piperidin-1-yl)butanes

Paul E. Finke,^{a,*} Laura C. Meurer,^a Bryan Oates,^a Sander G. Mills,^a Malcolm MacCoss,^a Lorraine Malkowitz,^b Martin S. Springer,^b Bruce L. Daugherty,^b Sandra L. Gould,^b Julie A. DeMartino,^b Salvatore J. Siciliano,^b Anthony Carella,^b Gwen Carver,^c Karen Holmes,^c Renee Danzeisen,^c Daria Hazuda,^c Joseph Kessler,^c Janet Lineberger,^c Michael Miller,^c William A. Schleif^c and Emilio A. Emini^c

^aDepartment of Medicinal Chemistry, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA ^bDepartment of Immunology Research, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA ^cDepartment of Antiviral Research, Merck Research Laboratories, PO Box 4, West Point, PA 19486, USA

Received 18 September 2000; accepted 9 November 2000

Abstract—(2*S*)-2-(3,4-Dichlorophenyl)-1-[*N*-(methyl)-*N*-(phenylsulfonyl)amino]-4-[spiro(2,3-dihydrobenzthiophene-3,4'-piperidin-1'-yl)]butane *S*-oxide (3) has been identified as a potent CCR5 antagonist lead structure having an $IC_{50} = 35$ nM. Herein, we describe the structure–activity relationship studies directed toward the requirement for and optimization of the C-2 phenyl fragment. The phenyl was found to be important for CCR5 antagonism and substitution was limited to small moieties at the 3-position (13 and 16: X = H, 3-F, 3-Cl, 3-Me). © 2001 Published by Elsevier Science Ltd.

Human immunodeficiency virus type-1 (HIV-1) is an enveloped virus that must fuse its envelope with the plasma membrane of its host cell to gain cell entry.¹ CCR5, a seven-transmembrane receptor for the βchemokines MIP-1α, MIP-1β, and RANTES,² has been identified as a primary co-receptor with CD4 for cell entry of macrophage-tropic (M-tropic or R5) HIV-1 isolates.³ Individuals homozygous for a 32-base pair deletion in the gene for CCR5 do not express functional receptor on their cell surfaces and have been identified as being highly resistant to HIV-1 infection, 4 while infected individuals heterozygous for the defective gene appear to exhibit delayed disease progression.⁵ Given the importance of CCR5 for the establishment, and possible maintenance, of HIV-1 infection in vivo, and the lack of an overt detrimental phenotype in humans that do not express functional CCR5, numerous efforts have been initiated in an effort to identify suitable CCR5 antagonists for use as potential anti-HIV-1 therapeutic agents. 6-9

Towards this end, there have now been several reports of CCR5 antagonists in the patent literature.^{1,10} To date, the most established structure is TAK-779 (1),^{11,12} which is actually a dual CCR5 and CCR2b antagonist having binding affinities of 1.4 and 27 nM, respectively.

In our first manuscript in this series, 13 the discovery of several CCR5 selective (2S)-1-(N-alkyl-N-arylsulfonylamino)-2-phenyl-4-(piperidin-1-yl)butane structures (2) were identified through screening of the Merck sample collection and our initial structure–activity relationships (SARs) pertaining to the C-1 N-alkyl-N-arylsulfonamide moiety of 2 were reported. This work resulted in the identification of (2S)-2-(3,4-dichlorophenyl)-1-[(N-methyl-N-phenylsulfonyl)amino]-4-[spiro(2,3-dihydrobenzthiophene-3,4'-piperidin-1'-yl)]butane S-oxide (3, mixture of R and S-sulfoxides) as our initial key lead structure having an $IC_{50} = 35 \, \text{nM}$ for inhibition of $[^{125}I]$ -MIP-1 α

^{*}Corresponding author. E-mail: paul finke@merck.com

binding to CCR5.¹⁴ Herein, the SAR for the central C-2-phenyl moiety will be described.

Using chemistry similar to that reported previously from these laboratories ^{13,15,16} (see Scheme 1) and several of the available intermediates, an initial SAR for the central phenyl ring was rapidly developed that indicated substitution on the phenyl could dramatically effect CCR5 inhibition, monosubstitution at the 3-position being preferred (see Table 1, compound 13d). In addition, some early work on other piperidine derivatives indicated that the 4-phenylpiperidine as in 4 could replace the spiropiperidine of 3 while retaining most of the CCR5 binding potency, but with a greatly simplified structure.

Thus, in order to more easily explore the central phenyl ring substitution, a modified synthetic approach (see Scheme 2) was developed which could introduce a substituted aryl group at C-2 late in the synthesis rather than in the starting material as required in our established route. This new approach utilized an appropriately difunctionalized vinyl—tin intermediate 20, which underwent Pd-catalyzed coupling¹⁷ with a variety of aryl bromides to give the substituted styrenes 22. Subsequent hydrogenation afforded the final racemic compounds in only two steps. The simplified piperidine moiety from above was employed in this alternate tin-coupling route.

The synthesis of **3** and several initial substituted C-2 phenyl derivatives (Scheme 1) began with the phenylacetic acids **5**. Alkylation of **5** with allyl bromide using lithium hexamethyldisilylamide (LiHMDS) provided racemic **6** that, if desired, could usually be resolved by repeated crystallization of the (S)-(-)- α -methylbenzylamine salts

to afford the desired chiral (2S) enantiomer **6a**. Use of (R)-(+)- α -methylbenzylamine afforded the (2R) enantiomer **6b**. Alternatively, chemical resolution was possible by conversion of **6** to the diastereomeric (S)-(-)-4-benzyl-2-oxazolidinone derivatives, separation on silica gel, and hydrolysis with lithium hydroxide/hydrogen peroxide in THF. Thus, the C-2 stereochemistry and phenyl substitution were set at the start. Conversion of **6a** to the amides **7** was done via the acid chlorides followed by treatment with methylamine. Reduction of **7** with DIBAL-H provided the *N*-methylamine intermediates **8**, which

Scheme 1. Reagents: (a) LiHMDS, THF, -70°C; (b) allyl bromide, -70°C to rt; (c) (*S*)-(-)-α-methylbenzylamine (0.6 equiv), *i*-PrOH, then two recrystallizations affords **6a**; (d) (*R*)-(+)-α-methylbenzylamine (1 equiv), *i*-PrOH, then two recrystallizations affords **6b**; (e) 2 N HCl, water, EtOAc (three extractions); (f) (Me₃CCO)₂O, TEA, THF, -70°C to rt, then lithium salt of (*S*)-4-benzyl-2-oxazolidine in THF, -70°C to rt; (g) LiOH, H₂O₂, 2:1 v/v THF/water, 0°C, then Na₂SO₃ (aq), rt; (h) oxalyl chloride, DMF (cat), DCM, rt; (i) MeNH₂ (40% aq, 5 equiv), THF, 0°C to rt; (j) DIBAL-H, THF, rt; (k) PhSO₂Cl, DIPEA, DCM, rt; (l) OsO₄ (cat), NMO, 2:1:1 v/v/v acetone/t-butanol/water, rt; (m) NaIO₄, 4:1 v/v THF/water, rt; (n) 11-HCl, DIPEA, NaBH(OAc)₃, DCE, rt; (o) Oxone[®] (1.2 equiv), MeOH, -20°C, 2-5 min; (p) Oxone[®] (3 equiv), MeOH, rt; (q) **15**, AcOH, NaBH(OAc)₃, DCE, rt.

were then sulfonylated with benzenesulfonyl chloride to afford the functionalized right hand portion (9). A two-step oxidation of the allyl group with osmium tetroxide/*N*-methylmorpholine-*N*-oxide (NMO) followed by sodium periodate cleavage of the intermediate diols afforded the

Table 1. Structures and CCR5 binding activities for the spiropiperidine derivatives **3**, **12**, **13**, and **14**

Compound	Structure		CCR5 ^a
	n	R	$IC_{50} (nM)^b$
12a	0	(S)-3,4-diCl-Phenyl	1000
3	1	(S)-3,4-diCl-Phenyl	35
14a	2	(S)-3,4-diCl-Phenyl	100
12b	0	(R/S)-Phenyl	450
13b	1	(R/S)-Phenyl	35
14b	2	(R/S)-Phenyl	30
13c	1	(R/S)-2-Cl-Phenyl	2000
14c	2	(R/S)-2-Cl-Phenyl	1300
12d	0	(S)-3-Cl-Phenyl	270
13d	1	(S)-3-Cl-Phenyl	10
14d	2	(S)-3-Cl-Phenyl	15
13e	1	(S)-4-Cl-Phenyl	270
13f	1	(S)-4-F-Phenyl	570
13g	1	(R/S)-3,5-diCl-Phenyl	90
14g	2	(R/S)-3,5-diCl-Phenyl	110
13h	1	(S)-3,4-OCH ₂ O-Phenyl	200
14h	2	(S)-3,4-OCH ₂ O-Phenyl	80
13i	1	(R/S)-2-Thienyl	180
14i	2	(R/S)-2-Thienyl	110
13j	1	(R/S)-3-Thienyl	100
14j	2	(R/S)-3-Thienyl	50
13k	1	(R/S)-Cyclohexyl	>1000 (36%)°
14k	2	(R/S)-Cyclohexyl	>1000 (42%)°

^aSee ref 13 for procedures.

aldehydes **10**. Reductive amination¹⁸ of the spiropiperidine **11** with **10** using sodium triacetoxyborohydride in dichloroethane (DCE) provided the sulfide derivatives **12a**–**k**. Final oxidation of the sulfur with 1 equiv of Oxone^{® 19} at -20 °C afforded the sulfoxides **3** and **13b**–**k** (as 1:1 mixtures of sulfoxide diastereomers), while oxidation with excess Oxone[®] at room temperature afforded the corresponding sulfones **14a**–**k** (see Table 1). Alternatively, reductive alkylation of 4-phenylpiperidine (**15**) with aldehydes **10** resulted in the corresponding substituted central phenyl derivatives **16a**–**k** (see Table 2).

Since the activities of the above 4-phenylpiperidine derivatives had only a 3-fold or less reduction in CCR5 binding affinity compared to the corresponding sulfoxide compounds (see below), this simplified left-hand moiety was utilized for further exploration of the central phenyl in the following modified synthetic route (Scheme 2). 2-Butyn-1,4-diol (17) was treated with 2.5 equiv of triphenylphosphine-dibromide to form the dibromide 18. In a one-pot double displacement sequence, first with 0.75 equiv of the preformed sodium salt of N-methylbenzenesulfonamide followed by excess 4-phenylpiperidine (15), the crude 18 afforded the disubstituted alkyne 19 in 42% yield. Hydrostannylation²⁰ of 19 was carried out with PdCl₂(Ph₃P)₂ in THF to give primarily the desired readily separable regioisomer 20 over 21 in a 4:1 ratio (55 and 17%, 32% recovered 19).21 The best conditions found for the coupling of 20 to a variety of substituted phenyl bromides and 2- and 3-bromopyridine utilized PdCl₂(Ph₃P)₂ in N-methylpyrrolidine (NMP) at 70 °C in the presence of potassium carbonate. The inclusion of potassium carbonate neutralized any acid formed and served to provide a more reactive ligand for the Pd. 17 Yields of the coupled styrene products 22 were between 20 and 40%. Hydrogenation of 22 to the final racemic products 16 was done with Pd(OH)₂ in methanol and required the addition of acetic acid. Unfortunately, these conditions also promoted the partial hydrogenolysis of the allylic piperidine to give 23 and resulted in lower yields of 16, usually about 50%. The

Scheme 2. Reagents: (a) (Ph₃P)-Br₂ (2.25 equiv), MeCN, 0°C to rt; (b) PhSO₂NHMe (0.75 equiv), NaH (60%), DMF, 0°C; (c) crude **18** in DMF, 0°C, addition of PhSO₂NMe-Na (0.75 equiv) from (b), 0°C, 2 h; then **15** (1.5 equiv), DIPEA, rt; (d) Bu₃SnH, PdCl₂(Ph₃P)₂ (2 mol%), THF, rt; (e) X-PhBr, PdCl₂(Ph₃P)₂ (3 mol%), K₂CO₃, N-methylpyrrolidine, 70°C, 16 h; (f) H₂ (50 psi), 20% Pd(OH)₂ (cat), HOAc, MeOH, rt.

^bThe IC_{50} results are an average of three independent titrations having calculated standard errors below 15%. The assay-to-assay variation was generally ± 2 -fold based on the results of the standard compound **13d**. ^cThese compounds gave the indicated % inhibition at $1000 \, \text{nM}$.

Table 2. Structures and CCR5 binding activities for the 4-phenyl-piperidine derivatives **16** and **27**

	g	CCP 50
0 1	Structure	CCR5a
Compound	R	$IC_{50} (nM)^b$
16b	(R/S)-Phenyl	120
16c	(R/S)-2-Cl-Phenyl	3000
16d	(S)-3-Cl-Phenyl	30
16f	(S)-4-F-Phenyl	~1000
16g	(R/S)-3,5-diCl-Phenyl	300
16h	(S)-3,4-OCH ₂ O-Phenyl	300
16i	(R/S)-2-Thienyl	400
16j	(R/S)-3-Thienyl	100
16k	(R/S)-Cyclohexyl	>1000 (26%) ^c
16l	(R/S)-3-F-Phenyl	100
16m	(R/S)-3-Me-Phenyl	80
16n	(R/S)-3-Et-Phenyl	110
16o	(R/S)-3-CF ₃ -Phenyl	500
16p	(R/S)-3-OMe-Phenyl	>1000 (45%) ^c
16q	(R/S)-3-Biphenyl	$\sim 10,000 (76\%)^{d}$
16r	(R/S)-4-Me-Phenyl	200
16s	(R/S)-4-OMe-Phenyl	>1000 (40%)°
16t	(R/S)-3,5-diMe-Phenyl	160
16u	(R/S)-3,4-diF-Phenyl	570
16v	(R/S)-3,4-diMe-Pphenyl	60
16w	(R/S)-3-Me,4-F-Phenyl	180
16x	(R/S)-3-F,4-Me-Phenyl	110
16y	(R/S)-2-Pyridyl	$\sim 10,000 (61\%)^{d}$
16z	(R/S)-3-Pyridyl	>10,000 (43%) ^d
16aa	(R/S)-2-Naphthyl	720
27a,b	(R/S) Benzyl	$\sim 10,000 (68\%)^{d}$

^aSee ref 13 for procedures.

^dThese compounds gave the indicated % inhibition at 10,000 nM.

final compounds **16l–aa** prepared by this route are also listed in Table 2.

In order to prepare the C-2 benzyl derivative, the reaction of **20** with benzyl bromide was attempted. However, alkylation of the piperidine nitrogen occurred to give the quaternary amine. Thus, a modified route was utilized starting from the diol **17** (Scheme 3). Using analogous chemistry as above, the vinyl stannane **24** afforded 2-benzylbut-2-ene-1,4-diol (**25**). The incorporation of the sulfonamide and piperidine moieties was again achieved in a single reaction via the dibromide to afford **26a,b**. Final hydrogenation then afforded **27a,b** as a mixture of racemic regioisomers.

Finally, use of the isomeric stannane 21 afforded the isomerically pure racemic C-3 phenyl derivative 28 and hydrogenation of the alkyne 19 yielded the des-phenyl derivative 29 (Scheme 4).

These compounds were then evaluated for CCR5 affinity utilizing a [125 I]-MIP-1 α binding assay. 14 An initial survey of monosubstitution at the 2-, 3-, and 4-phenyl

Scheme 3. Reagents: (a) Bu₃SnH, PdCl₂(Ph₃P)₂ (2 mol%), THF, rt; (b) benzyl bromide, PdCl₂(Ph₃P)₂ (3 mol%), K_2CO_3 , NMP, $70^{\circ}C$, 6 h; (c) (Ph₃P)-Br₂ (2.25 equiv), MeCN, $0^{\circ}C$ to rt; (d) PhSO₂NHMe (0.75 equiv), NaH (60%), DMF, $0^{\circ}C$; (e) crude dibromide in DMF, $0^{\circ}C$, addition of PhSO₂NMe-Na (0.75 equiv) from (d), $0^{\circ}C$, 2 h; then 15 (1.5 equiv), DIPEA, rt; (f) H₂ (50 psi), 20% Pd(OH)₂, HOAc, MeOH, rt.

Scheme 4.

positions resulted in a clear preference for substitution at the 3-position as seen in the chloro sulfoxide series 13c, **13d**, and **13e** (IC₅₀ = 2000, 10 and 270 nM, respectively). In agreement with previous results with 12a, 3, and 14a, 13 in all cases the sulfides had the poorest CCR5 binding affinity (i.e. 12b and d, $IC_{50} = 450$ and 270 nM) while the sulfone derivatives usually resulted in only slightly poorer binding than the corresponding sulfoxides (compare 13c and 14c, 10 and 15 nM, respectively). Thus, the following discussion deals mostly with the sulfoxide results. While it would have been of interest, any possible sulfoxide stereochemical preference for CCR5 interaction was not determined since the sulfoxide isomers were not separable. In agreement with the improved CCR5 binding seen with 13d and 14d, enhanced antiviral activity was also observed (see below).

Removal of both of the chlorines as in 13b $(IC_{50} = 35 \text{ nM}, \text{ racemic}^{22})$ actually resulted in a possible 2-fold improvement in binding compared to the 3,4dichloro lead compound 3 ($IC_{50} = 35 \text{ nM}$, chiral²²) and within a 2-fold reduction compared to 13d. Dichloro substitution at the 3,5-positions (13g, $IC_{50} = 90 \,\text{nM}$, racemic) was not additive and actually showed a moderate loss in potency compared to 13d. 3,4-Disubstitution as in the methylenedioxy derivative 13h (IC₅₀ = 200 nM, chiral) decreased affinity relative to 3. Replacement of the phenyl with either a 2- or 3-thienyl (13i and 13j, 180 and 100 nM, both racemic) led to a moderate loss in binding while the cyclohexane analogue 13k resulted in very poor activity. Thus, from this initial survey of substitutions on the C-2 phenyl, limited substitution at the 3-position or even no substitution appeared to be preferred.

^bThe IC_{50} results are an average of three independent titrations having calculated standard errors below 15%. The assay-to-assay variation was generally ± 2 -fold based on the results of the standard compound **13d**. ^cThese compounds gave the indicated % inhibition at 1000 nM.

From our initial screening results, there had been hints that the spiro-piperidine was not required and that a simple 4-phenylpiperidine might have similar binding (data not shown). Indeed, this modification in the above cases afforded very similar results, being within a factor of three in all cases as exemplified with the best 3-chloro compounds 13d and 16d having IC₅₀s of 10 versus 30 nM. Also, the unsubstituted compound 16b was only 2-fold lower in binding than **16d**, the best 3-substituted compound in this series ($IC_{50} = 120 \text{ nM}$, racemic, versus 30 nM chiral). Thus, this simplified piperidine derivative was utilized in a more rigorous exploration of the phenyl substitution. From these additional derivatives, other small substituted analogues, such as 3-fluoro (161, $IC_{50} = 100 \text{ nM}$, racemic), 3-methyl (16m, $IC_{50} = 80 \text{ nM}$, racemic), and even 3-ethyl (16n, $IC_{50} = 110 \text{ nM}$, racemic), exhibited potency within 2-fold of 16d. Larger groups, such as trifluoromethyl (160), methoxy (16p), and phenyl (16q), all had diminished CCR5 affinity. Surprisingly, the binding of the 4-methyl compound 16r $(IC_{50} = 200 \text{ nM}, \text{ racemic})$ was appreciably better than would be expected from the 4-Cl (13e) and 4-F (13f and 16f) results, although the 4-MeO compound (16s) was inactive. The benign effect of the 4-methyl carried over to the 3,4-dimethyl derivative (16v, $IC_{50} = 60 \text{ nM}$, racemic), which appeared to be equipotent with the 3-chloro 16d, as well as the 3-F,4-Me compound 16x (IC₅₀ = 110 nM, racemic). However, other 3,4-disubstitution was generally detrimental as anticipated (see 16u and 16w). Replacement of the central phenyl ring with a 2- or 3-pyridyl (16y and 16z) resulted in substantial loss of CCR5 activity. Extension of the phenyl by a methylene unit was not tolerated as seen with the benzyl derivatives 27a,b for which there was no apparent activity for any of the four isomers. All the vinyl intermediates 22 and the des-piperidine compound 23 were inactive (Scheme 2, IC₅₀>4000 nM, data not shown). The importance of the phenyl was also demonstrated by the lack of CCR5 affinity for the C-3 phenyl isomer 28 and the des-phenyl compounds **19** and **29** (61, 5, and 20% I at 10,000 nM, respectively).

The lead structure 3 was initially characterized in an isolated peripheral blood mononuclear cell (PBMC) viral replication assay²³ using the R5-tropic HIV-1 YU-2 isolate and gave IC₉₅ values of 6 to 12 µM.¹³ This activity indicated that inhibition of viral entry was possible with these small molecule CCR5 antagonists. With the enhancement in the CCR5 binding assay obtained with the 3-chlorophenyl derivative 13d in the spiro-piperidine series, better results were also seen in this antiviral assay, giving an IC₉₅ as low as 1500 nM,²⁴ while the sulfone **14d** afforded an IC₉₅ of 3000 nM in the same assay. Use of the different R5-tropic strain SF162 afforded IC₉₅'s of 800 and 6000 nM for 13d and 14d, respectively. The unsubstituted compound 13b also afforded inhibition similar to 13d in the same assay. The best thiophene derivative (14j, $IC_{50} = 50 \text{ nM}$) was not active in this assay (IC₉₅>25,000 nM). As expected from the CCR5 binding data, the antiviral activity was poorer in the 4-phenylpiperidine series, again the best activity was seen with the 3-chloro derivative **16d**, as well as the 3methyl 16m, both having IC₉₅ of 6000 nM against the

YU-2 strain. As expected, **13d** failed to give any inhibition when the X4-tropic NL4-3 strain was used (data not shown).

Herein, the results of our SAR study of the central phenyl ring of lead compound 3 were described. In the spiro-piperidine series, the 3-chloro derivative 13d was identified as a having a 4-fold improvement in CCR5 binding affinity and resulted in enhanced antiviral inhibition in a PBMC based assay with an IC95 as low as 1500 nM and served thereafter as a standard in this assay.24 Selectivity for CCR5 was maintained by 13d in that the IC₅₀'s for CCR1, CCR2, CCR3, and CXCR4 were all >10,000 nM. Both 13d and 14d showed modest pharmacokinetics in the rat at 1 mg/kg iv and 10 mg/kg oral ($t_{1/2}$ =0.7 and 0.8 h, F=3 and 2%, respectively). In addition, the unsubstituted compound 13b was found to be nearly as potent an antagonist as 13d and 14d. Comparable affinity was also found in the 4-phenylpiperidine series which allowed identification of other small substituents, such as 3-F (16i) and 3-Me (16m), as being nearly equipotent to 3-Cl. These results allowed the simplification of the synthesis of further piperidine derivatives as well as other structural modifications which will be reported in the future.

References and Notes

- 1. For a review of HIV-1 entry mechanisms and current inhibitors, see: Blair, W. S.; Lin, P.-F.; Meanwell, N. A.; Wallace, O. B. *Drug Discovery Today* **2000**, *5*, 183.
- 2. For a recent chemokine review, see Baggiolini, M.; Dewald, B.; Moser, B. *Annu. Rev. Immunol.* **1997**, *15*, 675.
- 3. For a review of the co-receptor search, see: Fauci, A. S. *Nature* **1996**, *384*, 529.
- 4. Liu, R.; Paxton, W. A.; Choe, S.; Ceradini, D.; Martin, S. R.; Horuk, R.; MacDonald, M. E.; Stuhlmann, H.; Koup, R. A.; Landau, N. R. *Cell* **1996**, *86*, 367.
- 5. Michael, N. L.; Chang, G.; Louie, L. G.; Mascola, J. R.; Dondero, D.; Birx, D. L.; Sheppard, H. W. *Nature Med.* 1997, 3 338
- 6. Bates, P. Cell 1996, 86, 1
- 7. Moore, J. P. Science 1997, 276, 51.
- 8. Cohen, O. J.; Kinter, A.; Fauci, A. S. Immunol. Rev. 1997, 159, 31.
- 9. D'Souza, M. P.; Cairns, J. S.; Plaeger, S. F. JAMA 2000, 284, 215.
- 10. Horuk, R.; Ng, H. P. Med. Res. Rev. 2000, 2, 155.
- 11. Shiraishi, M.; Aramaki, Y.; Seto, M.; Imoto, H.; Nishikawa, Y.; Kanzaki, N.; Okamoto, M.; Sawada, H.; Nishimura, O.; Baba, M.; Fujino, M. *J. Med. Chem.* **2000**, *43*, 2049. 12. Dragic, T.; Trkola, A.; Thompson, D. A. D.; Cormier, E. G.; Kajumo, F. A.; Maxwell, E.; Lin, S. W.; Ying, W.; Smith, S. O.; Sakmar, T. P.; Moore, J. P. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, *97*, 5639.
- 13. Dorn, C. P.; Finke, P. E.; Oates, B.; Budhu, R. J.; Mills, S. G.; MacCoss, M.; Malkowitz, L.; Springer, M. S.; Daughtery, B. L.; Gould, S. L.; DeMartino, J. A.; Siciliano, S. J.; Carella, A.; Carver, G.; Holmes, K.; Danzeisen, R.; Hazuda, D.; Kessler, J.; Lineberger, J.; Miller, M.; Schleif, W. A.; Emini, E. A. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 259.
- 14. For a description of the binding assay, see ref 13, footnote 25. The CCR5 binding titrations were usually done in triplicate with standard errors of <15%. Assay-to-assay variability was generally ± 2 -fold based on the standard compound 13d.

- 15. MacCoss, M.; Mills, S. G.; Shah, S. K.; Chiang, Y.-C. P.; Dunn, P. T.; Koyama, H. US Patent 6,013,652, 2000; *Chem. Abstr.* **2000**, *132*, 78470s.
- 16. Hale, J. J.; Finke, P. E.; MacCoss, M. Bioorg. Med. Chem. Lett. 1993, 3, 319.
- 17. For reviews of the Stille coupling, see: (a) Stille, J. K. *Angew. Chem., Int. Ed. Engl.* **1986**, *25*, 508; or (b) Stille, J. K. *Pure Appl. Chem.* **1985**, *57*, 1771.
- 18. Abdel-Magid, A. F.; Carson, K. G.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. *J. Org. Chem.* **1996**, *61*, 3849.
- Trost, B. M.; Curran, D. P. Tetrahedron Lett. 1981, 22, 1287.
 Zhang, H. X.; Guibe, F.; Balavoine, G. J. Org. Chem. 1990, 55, 1857.
- 21. NMR data for the isomeric stannanes **20** and **21**. Higher R_f product **20**: 1 H NMR (400 MHz, CDCl₃) δ 0.8–0.9 (m, 6H), 0.87 (t, J=7 Hz, 9H), 1.25–1.35 (m, 6H), 1.35–1.5 (m, 6H), 1.6–1.85 (2 m, 4H), 1.94 (dt, J=3 and 8 Hz, 2H), 2.43 (m, 1H), 2.69 (s, 3H), 3.01 (brt, J_{H-Sn} =25 Hz, 2H), 3.78 (d, J=6 Hz, 2H), 5.44 (brt, J=1.5 and 6 Hz, J_{H-Sn} =34 Hz, 1H), 7.1–7.3 (2 m, 5H), 7.5–7.65 (m, 3H), 7.77 (dd, J=1.5 and 7.0 Hz, 2H).
- Lower R_f product **21**: ¹H NMR (400 MHz, CDCl₃) δ 0.87 (t, J=7 Hz, 9H), 0.95–1.05 (m, 6H), 1.25–1.4 (m, 6H), 1.45–1.6 (m, 6H), 1.7–1.85 (m, 4H), 1.92 (dt, J=3 and 8 Hz, 2H), 2.42 (m, 1H), 2.48 (s, 3H), 2.96 (m, 2H), 3.74 (brt, J_{H-Sn} =23 Hz, 2H), 5.88 (brt, J=6 Hz, J_{H-Sn} =32 Hz, 1H), 7.1–7.3 (2 m, 5H), 7.5–7.65 (m, 3H), 7.77 (dd, J=1.5 and 7.0 Hz, 2H).
- 22. Since the C-2 (R) configuration was much less potent than the (S), ¹³ for comparison with chiral derivatives the effective IC₅₀ activities for the racemic derivatives can be assumed to be essentially half that indicated in the tables.
- 23. Condra, J. H.; Schleif, W. A.; Blahy, O. M.; Gabryelski, L. J.; Graham, D. J.; Quintero, J. C.; Rhodes, A.; Robbins, H. L.; Roth, E.; Shivaprakash, M.; Titus, D.; Yang, T.; Teppler, H.; Squires, K. E.; Deutsch, P. J.; Emini, E. A. *Nature* 1995, 374, 569.
- 24. There was extensive variation in the results from this PBMC assay as seen in the results for 13d, which varied from 1500 to 12,500 nM. Thus, 13d was always used as a standard and the results for the other compounds are reported in relation to the result for 13d in the same assay.